

Identifying patient-specific neopeptides for cell-based and vaccine immunotherapy across breast cancer classifications reveals rarely shared recurrent neopeptides

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Background

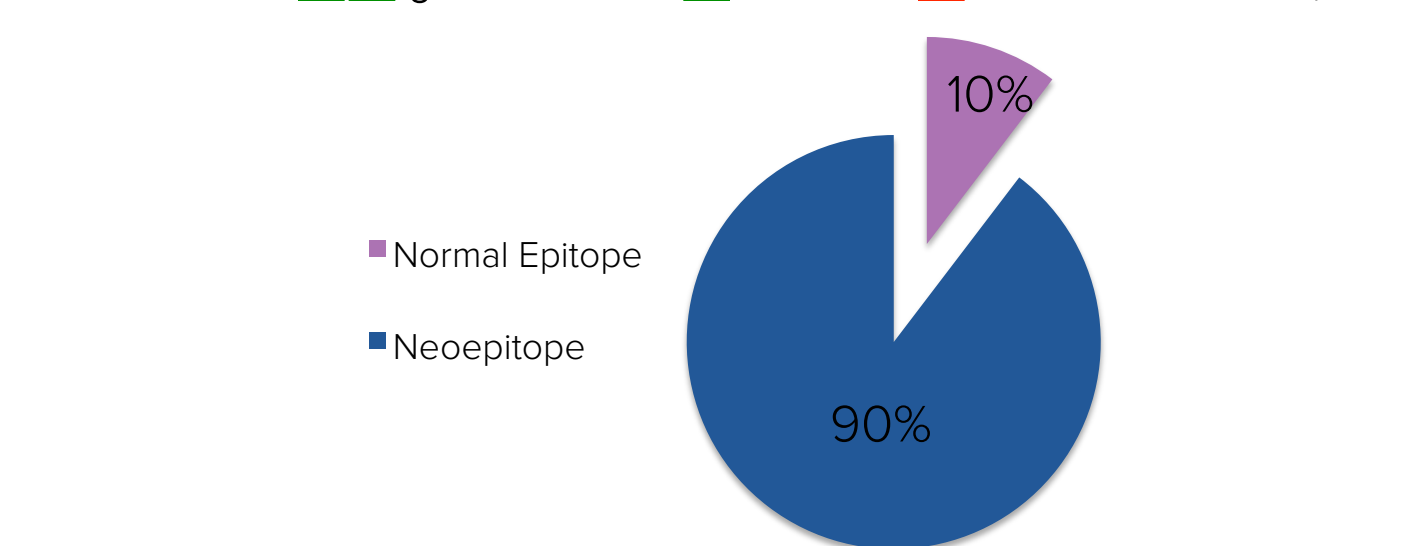
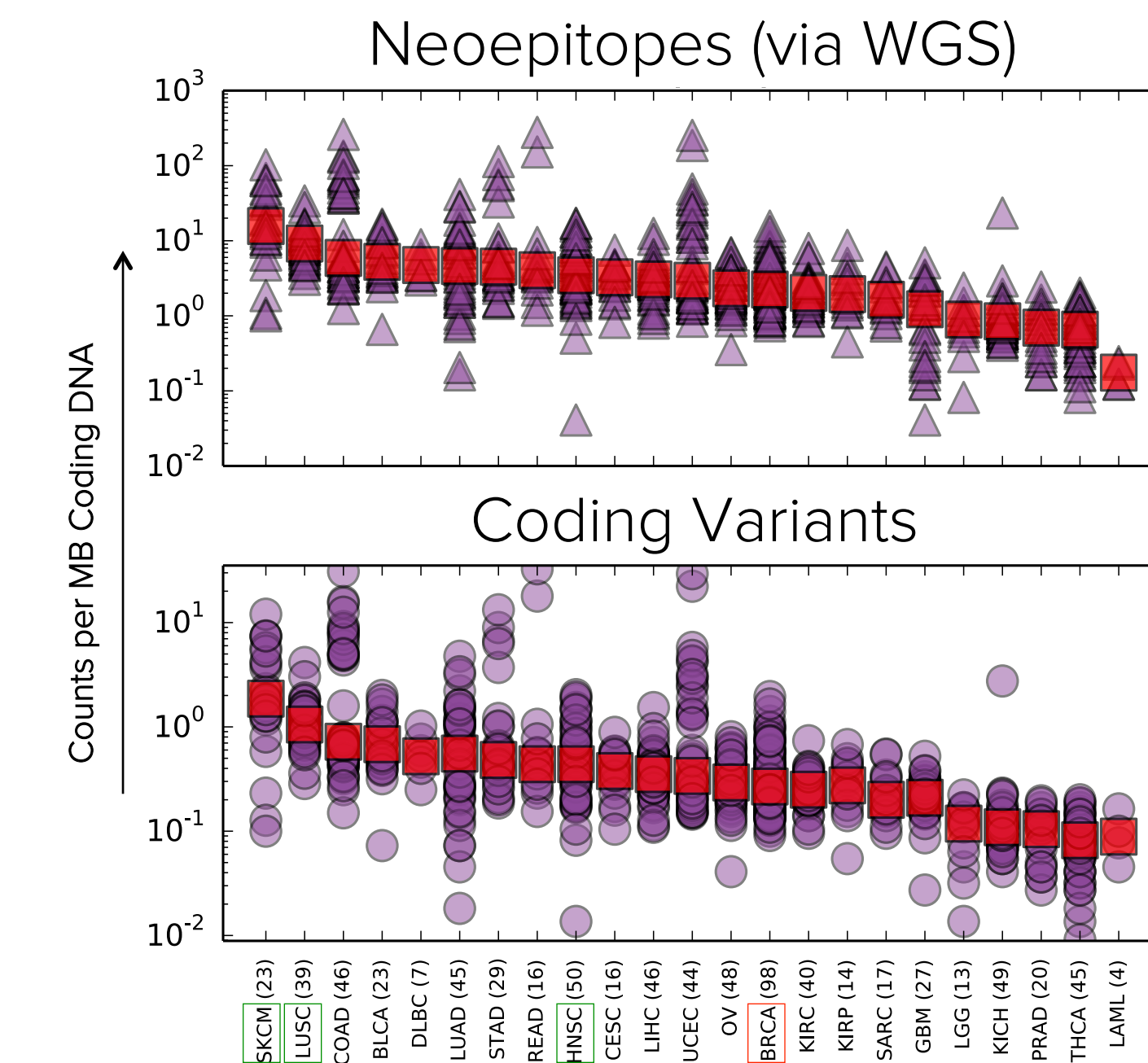
- Immunotherapies such as checkpoint inhibitors, CAR T cells, NK cells, and therapeutic vaccines are revolutionizing cancer medicine with remarkable responses in some patients.
- Several cancers have FDA approval for the use of check point inhibitors. And publications have suggested that mutational burden and neopeptide burden predict response to check point inhibitors.
- Cancers such as TNBC have few treatment options and would greatly benefit from novel immunotherapy approaches.
- We analyzed whole genome sequencing (WGS) and RNA sequencing (RNAseq) data from The Cancer Genome Atlas (TCGA) to identify neopeptides (tumor-specific antigens derived from somatic tumor mutations) that could be exploited to develop next-generation, patient-specific cancer immunotherapies.

Methods

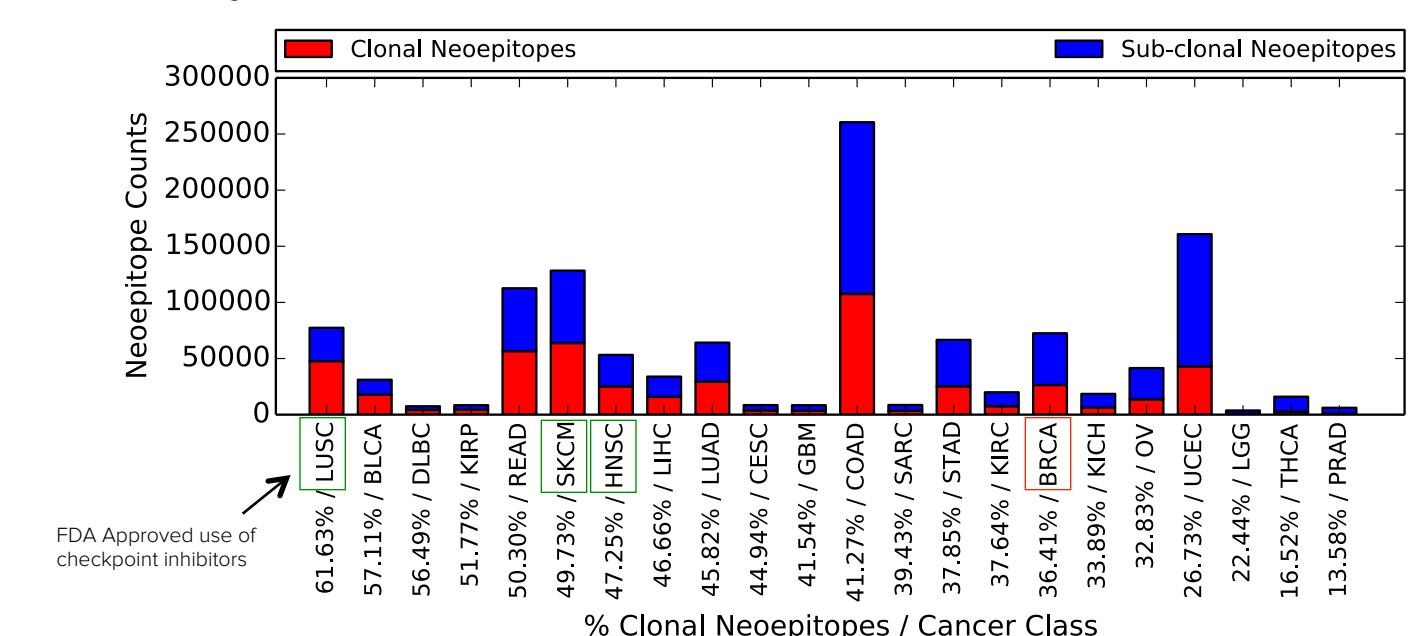
- TCGA WGS and RNASeq data were obtained from the University of California, Santa Cruz (UCSC) Cancer Genomics Hub (<https://cghub.ucsc.edu/>).
- Neopeptides were identified by creating all possible permutations of either 9-mer or 15-mer amino acid strings derived from somatic single nucleotide variants (SNVs) or insertions/deletions (indels) in coding regions.
- Potential neopeptides were filtered against all possible 9-mer and 15-mer sequences from reference human coding genes, in addition to all possible variation in dbSNP (<http://www.ncbi.nlm.nih.gov/SNP>) sites.
- In-silico HLA typing was performed using WGS and RNAseq data by alignment to the IMGT/HLA database. Typing results were obtained for HLA-A, HLA-B, HLA-C, and HLA-DRB1.
- NetMHC 3.4 (<http://www.cbs.dtu.dk/services/NetMHC-3.4/>) was used to predict MHC to neopeptide binding affinities.

Results

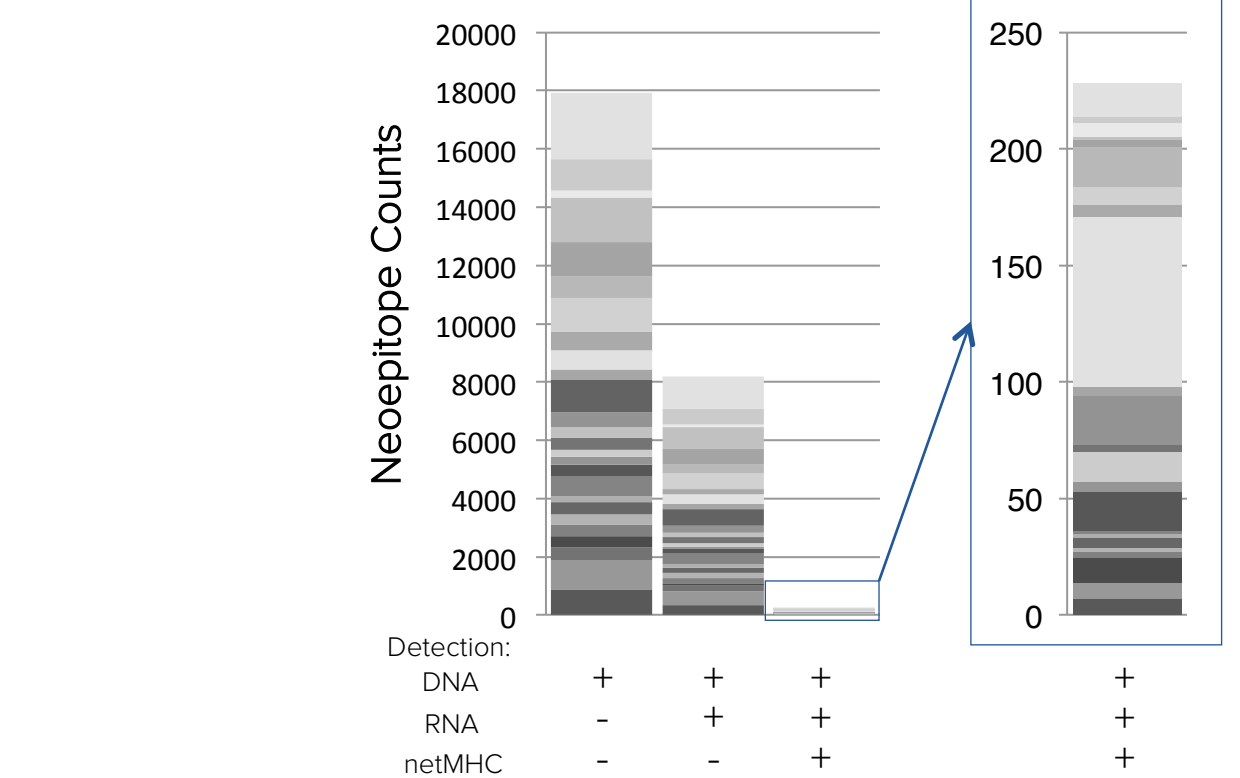
Cancer Neopeptide Loads Across TCGA Dataset



Clonality of Neopeptides across cancer classifications

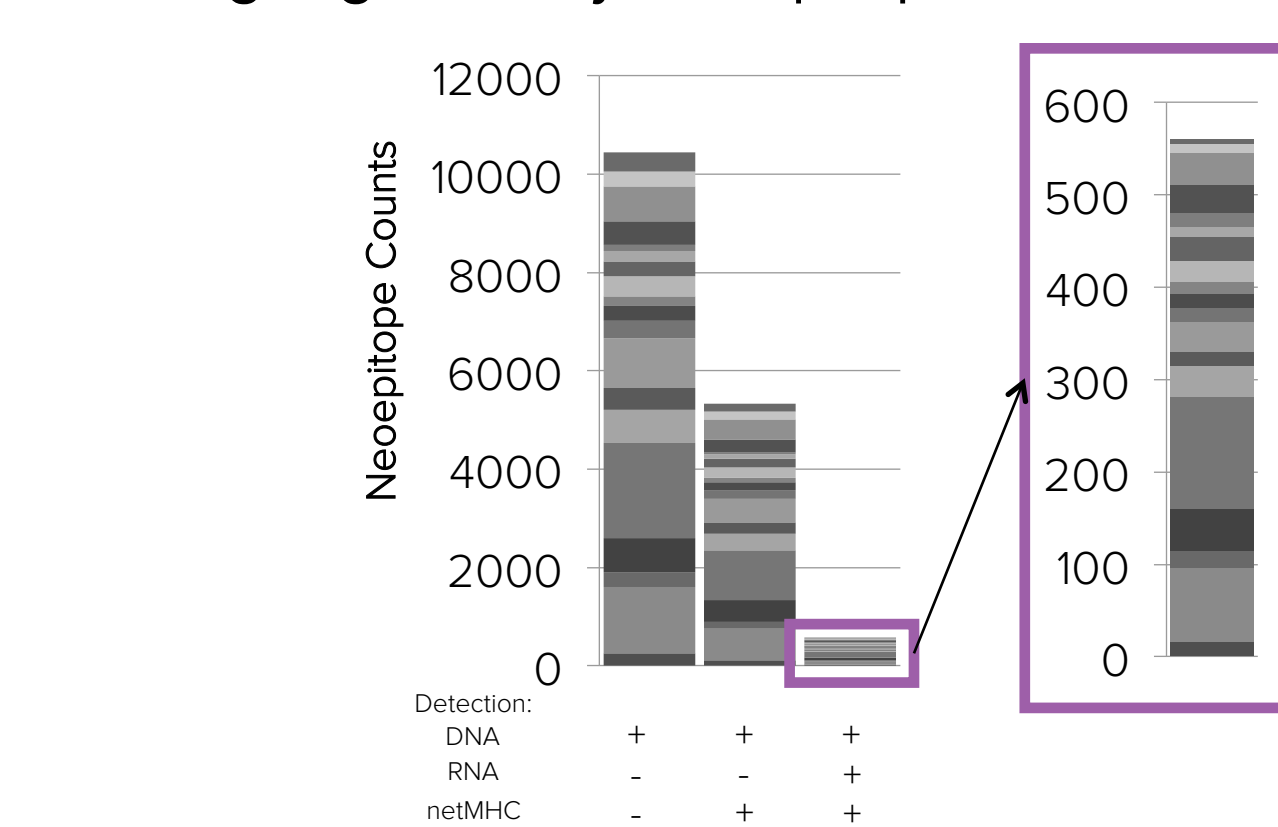


Filtering High Quality Neopeptides in TNBC



TCGA Barcode	UCSC id	HUGO Gene	TPM	Neopeptide	Protein Change	Normal	Bound Allele	Bind Strength
TCGA-E2-A14X	uc003ean.2	NAA50	229.85	PTDAHVLQK	p.A145T	PADAHVLQK	A*11:01	146nM
TCGA-E2-A1LL	uc001asj.3	FBXO2	187.36	LLLHLVLAAL	p.R57H	LLLRVLAAAL	A*02:01	18nM

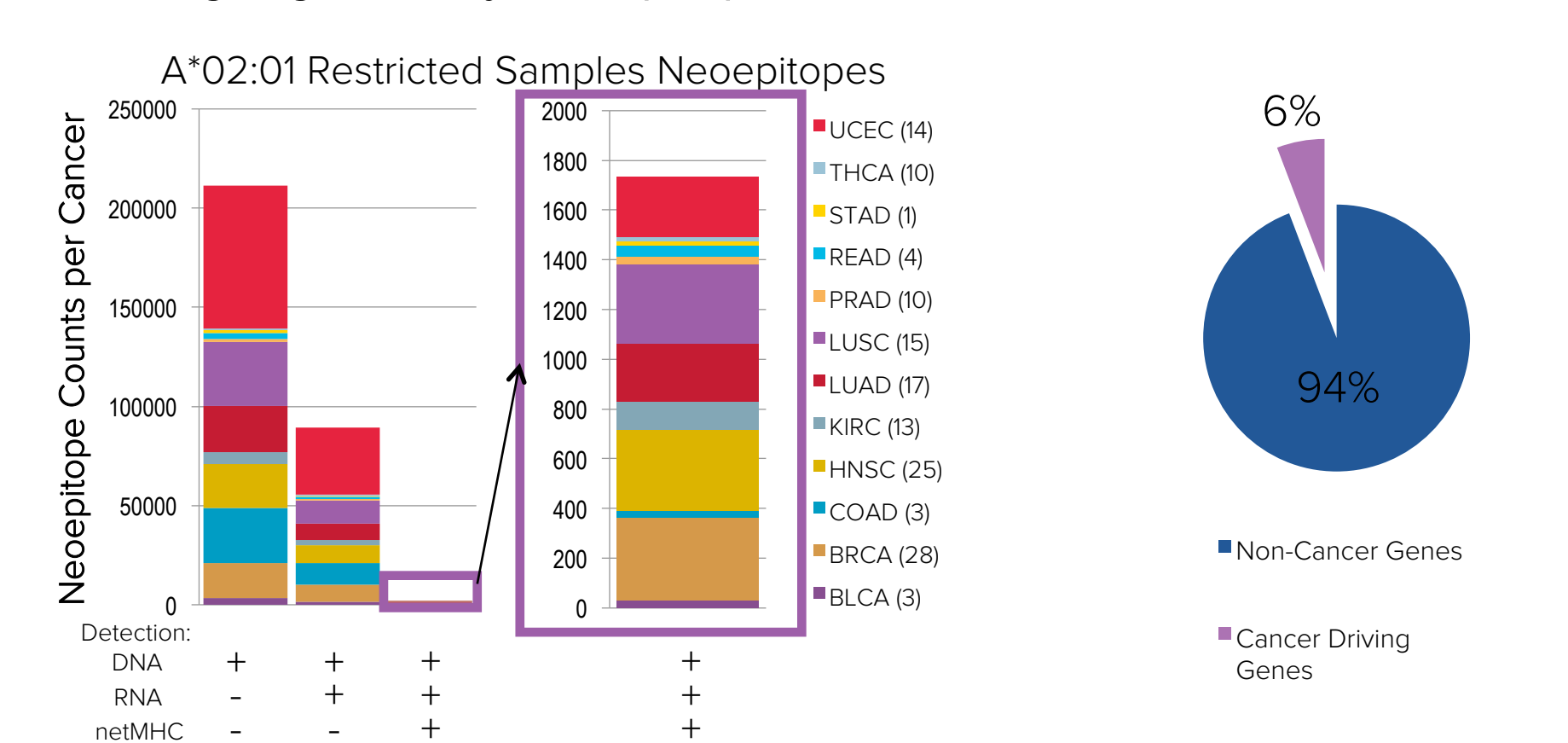
Filtering High-Quality Neopeptides in HER2+ BRCA



A Single Recurrent Neopeptide in TCGA HER2+ BRCA

TCGA Barcode	UCSC id	HUGO Gene	TPM	Neopeptide	Protein Change	Normal	Bound Allele	Bind Strength
TCGA-BH-A18R	uc003ean.2	FANCD2	21.39	FAKDGGGLVT	P714L	FAKDGGPVT	C*03:03	131nM
TCGA-AO-A0JM			14.12	FAKDGGGLVT	P714L	FAKDGGPVT	C*05:01	851nM

Filtering High-Quality Neopeptides Across Cancers



Shared Neopeptides Across Cancers

TCGA Barcode	UCSC id	HUGO Gene	Neopeptide	Protein Change	Normal	Cancers
TCGA-E2-A109, TCGA-CR-5249, TCGA-BA-6872, TCGA-CN-6989	uc001wxt.2	SOS2	YIHTHTFYV	p.T390I	YTHHTHTFYV	(3) HNSC, BRCA
TCGA-EW-A1J5, TCGA-21-1082, TCGA-GD-A2C5, TCGA-75-5147	uc001zyl.4	USP8	SQIWNLPV	p.R763W	SQIRNLPV	LUAD, BLCA, LUSC, BRCA

Conclusions

- Nearly all identified neopeptides are patient-specific. TNBC samples do not share any common neopeptides.
- Neopeptide-MHC interactions restrict more commonly shared mutations.
- Development of personalized immunotherapies is dependent on accurate DNA and RNA sequencing.
- Checkpoint inhibitor markers such as neopeptide load or PD1 expression will need to be screened before suggesting use of immunotherapy for breast cancer patients.

Acknowledgement

We would like to thank Kathryn Boorer, PhD, of NantHealth, LLC for writing assistance.

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